

ADVANCED ENZYME TECHNOLOGY

PROTEIN ENGINEERING (PART 1)

by

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by Main Author's Name

<http://ocw.ump.edu.my/course/view.php?id=602>

Chapter Description

- **Expected Outcomes**

- Describe about the general principal of protein engineering
- Discuss about the strategies of protein engineering
- Identify the differences between each strategies



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CONTENT

- General introduction
- Strategies of protein engineering
 - *De novo* design
 - Rational design
 - Site directed mutagenesis
 - Overlap extension PCR mutagenesis
 - Directed evolution
 - Random mutagenesis
 - Error-prone PCR



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Introduction

- ❖ Commonly, enzymes usage in chemical industries and other industrial applications are applied when **extremely specific catalysis are required**.
- ❖ However , in general enzymes are usually:
 - **Restricted to the amount of reactions they are able to catalyse**
 - **Reduced stability in both organic solvents and at elevated temperatures.**
- ❖ Therefore, **protein engineering** become a dynamic research area which consist of **efforts to design new enzymes with novel properties**, either through rational design or in vitro evolution.

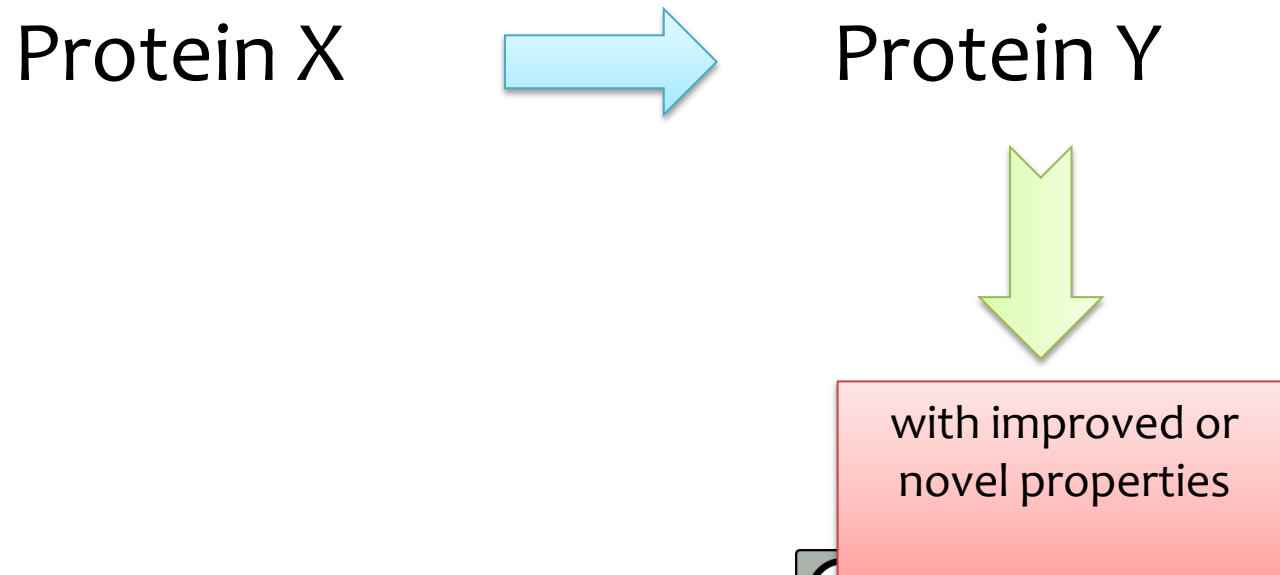


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- Protein Engineering : **Design** and **construct** proteins with **desired function(s)**



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Strategies of protein engineering

De novo design

Rational design

Directed evolution



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De novo design

“Knowledge-based protein design”



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- **enzymes with novel catalytic activities** are **still required** to be applied in numerous industrial biocatalytic processes.
- The ultimate goal of de novo protein design is to **develop catalytic activity** such that a biocatalyst can be readily obtained for any given chemical transformation.
- A **extremely challenging approach**, offering the broadest possibility for new structure.
- Aim: to discover the best amino acid sequence to guarantee folding in a selected structure (α -helical bundle or α/β -barrel)



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- **EXAMPLE:**

- ✓ **Methionine**: prefers rigid segments, the central part of helical segments, and buried regions of natural proteins
- ✓ **Threonine**: prefers flexible segments
- ✓ **Lysine**: a good helix former and prefers at the C-terminal, etc.
- ✓ **Leucine**: stabilizes helices, prefers buried regions, and is usually found in the middle positions in a helix
- ✓ **Glutamine**: hydrophilic residue; favourable ion pairs along one side of the helix to stabilize helix formation
- ✓ **Proline**: helix termination
- ✓ **Arginine**: stimulate a reversal in the overall peptide chain direction



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Rational Design

- Become the **earliest technique** of protein engineering
- Broadly applied to **introduce required properties into a protein of interest.**
- The **technique hinges on relating structure to function**, commonly through molecular modelling approach- Structure-function relationship of target protein.
- Advancements in rational design rely on **improvement in elucidation of structure, enhanced modelling procedures, and substantial new understanding into structure-function interactions**



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- Generally, proteins can be designed by **creating calculated variations** on a solved structure of protein with its sequence.
- **Create predictions of protein-sequence** that will fold to specific structures.
- **Function of protein is highly related on its structure** thus rational design approach utilize this association to develop function by designing proteins that contain a target structure or fold.



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- As a consequence, the **target structure or collective of structures must be identified** earlier in rational protein design approach.
- This is different to the other approach of protein engineering, for example directed evolution, where the structure information is not necessary and a various techniques are employed to identified proteins that attain a specific function.
- After that, the predicted sequences generated can be **tested experimentally**.



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Hypothesis : original sequence can be revised to optimized a desired function of protein

Which suggests that:

- Protein is **NOT** at an optimum for that function
- Sequence changes **without disruption of the structure** (otherwise it would not fold)
- New sequence designed is not **excessively dissimilar** from the original sequence or else lead to protein function loss.



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- There are several reasons to create specific DNA modifications
 - To **investigate changes in activity of protein** that happen as a consequence of DNA manipulation.
 - To **choose or screen for mutations** (at the DNA, RNA or protein level) that contain a target characteristic.
 - to be tuned to fit into the industrial marketplace.



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- Technique for rational design:

“Site-directed mutagenesis”



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Site-directed mutagenesis

- a technique to **design accurate and desired changes** in double stranded DNA of plasmid.
- The most **prevailing and broadly utilized** rational design approach, which **involves the accurate modification of specific residue(s)** in a target protein at the DNA level.
- These residue(s) are **usually recognized from the three-dimensional protein structure** (NMR or x-ray crystallography)



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- **If the target protein structure is not available,**
 - the available 3D structure of a homologous protein showing high sequence identity with the protein of interest can be used to build **a structural model of the target protein (homology modelling programs).**
- **If above options are NOT achievable,**
 - **sequence analysis programs for example NCBI BLAST can be employed to recognize the conserved residues** within a group of closely related proteins, thus suggested to be functionally important.



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- Three type of site-directed mutagenesis:

Insertions

- gaining one additional nucleotide

Deletions

- loss of one nucleotide

Substitutions

- change of one nucleotide (i.e. A → C)



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- Several PCR-based mutagenesis methods have been established and one of the most extensively employed methods is the **“overlap extension PCR mutagenesis”** method.



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Overlap extension PCR mutagenesis

- This strategies required two pairs of primer,
 - For each pair, one primer will comprises the mutant codon with a mismatched sequence.
- Then, all four primers (2 pairs) will be used in the first polymerase chain reaction (PCR) step,
 - in which two PCRs will take place, and two double-stranded DNA products will be produced.
- Upon denaturation and annealing of PCR amplification, two heteroduplexes are will be formed,
 - each strand of the heteroduplex involves the desired mutagenic codon.
- During second PCR step, DNA polymerase is then utilized to complete the overlapping 3' and 5' ends of each heteroduplex and the non-mutated primer set will be used to amplify the mutagenic product.



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